

XII. Survey of Plants for Steroidal Sapogenins and Other Constituents

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This is a report of the chemical examination of the second 1,000 samples received in a survey of plants for steroidal sapogenins. Data are given for 997 samples, representing 598 identified species, 201 unidentified lots, 392 genera, and 129 families. There is no previously published chemical information on about 60 per cent of the species examined. Quantitative data are given for the occurrence of 11 steroidal sapogenins. These were found almost exclusively in *Yucca*, *Agave*, and *Dioscorea*. A new sapogenin, named markogenin, was found in *Y. schidigera*. Qualitative data are given for the occurrence of the following groups of constituents: flavonols, alkaloids, tannins, and unsaturated sterols.

PAPER NUMBER VII in this series covered the survey of the first 1,000 samples (1). The present report is a continuation of it and covers the second 1,000 samples. The objective and assay procedures were described in detail in the former report and are not repeated here.

The data include the kinds and amounts of steroidal sapogenins found, and qualitative tests for saponins, flavonols, alkaloids, tannins, and unsaturated sterols. There are 997 samples, representing 598 identified species, 201 unidentified lots, 392 genera, and 129 families. There is no published chemical information on about 60 per cent of the species examined.

PROCUREMENT

The material represents a general sampling of the plant kingdom, as evidenced by the 364 genera in 107 families not included in the first thousand. Besides this continued general survey of the plant kingdom, an extensive collection was made of species in the genera *Dioscorea* and *Agave*.

Collections of *Agave* were obtained primarily by H. S. Gentry in Mexico and the southwestern United States and from the living collection maintained at the Huntington Botanical Garden in San Marino, Calif. He also collected numerous samples of *Dioscorea* in Mexico.

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In order to expedite the examination of such a large number of plant materials, groups of Laboratory workers were organized, each group handling certain phases of the whole procedure. The authors gratefully acknowledge the work of J. W. Garvin, Walter Rumph, R. A. Pierce, H. M. Neilson, G. H. Eppley, Theodore Perlestein, H. E. Kenney, Arthur Finchler, H. W. Jones, M. L. McClellan, Samuel Serota, R. F. Mininger, H. I. Sinnamon, A. E. Jones, C. S. Fenske, M. K. Scott, M. A. Morris, and J. R. Necho.

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General collections were obtained not only from the above-mentioned areas but also from several cooperators in the southeastern United States, Brazil, Colombia, Okinawa, and elsewhere. Approximately half the collections, however, were procured from permanent stock maintained at the Division of Plant Exploration and Introduction gardens at Coconut Grove, Fla.; Glenndale, Md.; and Chico, Calif.

RESULTS

A table has been prepared giving the data for each sample—its origin and identification, results of the hemolysis tests, kinds and amounts of steroidal sapogenins found, and qualitative findings for flavonols, alkaloids, tannins, and sterols. Because of space limitations, this table cannot be given here. It has, however, been prepared in processed form as AIC-367 and may be obtained on request from the U. S. Department of Agriculture, Eastern Regional Research Laboratory, Philadelphia 18, Pa.

In Table I the steroid data are rearranged to show their occurrence by species.

A positive hemolysis test for saponin was obtained in about 50% of the species. Steroidal sapogenins were found, however, almost exclusively in *Agave*, *Yucca*, and *Dioscorea*, the exceptions being *Manfreda* (Amaryllidaceae) and *Cordyline* (Liliaceae).

Four rarer steroidal sapogenins—kammogenin, samogenin, yammogenin, and yuccagenin—found in the first series were not found in the present one. However, a hitherto unknown one was found in the by-product leaf powder from the production of fiber from *Yucca schidigera* (sample 1686-0). It was named markogenin (2), spirostan-2 ξ ,3 β -diol, a 2,3-dihydroxy analog of sarsapogenin.

As stated in the previous report, the nature of the sapogenins found in plants is largely species specific. *Agave* species for the most part produce compounds with a *trans* ring A/B and with *iso* ring configuration at carbon 22. Depending on a number of environmental factors, tigogenin, hecogenin, manogenin, gitogenin, and occasionally chlorogenin are found singly or in combination. In this genus,

TABLE I.—OCCURRENCE OF THE VARIOUS SAPOGENINS BY SPECIES

Species	No. of Samples	Genin Content, (M. F. B., %)		Species	No. of Samples	Genin Content, (M. F. B., %)	
		Min.	Max.			Min.	Max.
Chlorogenin				<i>Agave virginica</i>	1	..	0.2
<i>Agave brandegeei</i>	1	..	Trace	<i>Agave sp.</i>	2	..	Trace
<i>Agave schidigera</i>	1	..	Trace	Markogenin			
<i>Agave sp.</i>	1	..	0.1	<i>Yucca schidigera</i>	1	..	0.2
<i>Manfreda sp.</i>	1	..	0.5	Sarsasapogenin			
<i>Yucca angustissima</i>	1	..	Trace	<i>Cordyline neo-caledonica</i>	1	..	0.5
9-Dehydrohecogenin				<i>Dioscorea bartlettii</i>	1	..	Trace
<i>Agave aurea</i>	1	..	0.05	<i>Yucca angustissima</i>	1	..	0.2
<i>Agave cerulata</i>	2	..	Trace	<i>Yucca arizonica</i>	1	..	0.3
<i>Agave nelsonii</i>	5	Trace	0.2	<i>Yucca baccata</i>	2	0.5	0.8
9-Dehydromanogenin				<i>Yucca elata</i>	2	0.5	1.3
<i>Agave aurea</i>	1	..	Trace	<i>Yucca faxoniana</i>	1	..	0.2
<i>Agave cerulata</i>	2	Trace	0.5	<i>Yucca schidigera</i>	1	..	2.9
<i>Agave goldmaniana</i>	1	..	Trace	<i>Yucca sp.</i>	2	0.8	0.9
<i>Agave nelsonii</i>	5	Trace	0.5	Smilagenin			
<i>Agave promontorii</i>	1	..	Trace	<i>Agave carchariodonta</i>	1	..	0.4
<i>Agave vexans</i>	1	..	Trace	<i>Agave difformis</i>	1	..	0.4
<i>Agave virginica</i>	1	..	0.1	<i>Agave goldmaniana</i>	1	..	0.3
Diosgenin				<i>Agave lophantha</i>	1	..	1.0
<i>Dioscorea bartlettii</i>	1	..	Trace	<i>Agave sp.</i>	1	..	1.0
<i>Dioscorea composita</i>	1	..	3.8	<i>Yucca sp.</i>	1	..	1.5
<i>Dioscorea glauca</i>	1	..	0.2	Tigogenin			
<i>Dioscorea villosa</i>	5	0.5	1.2	<i>Agave asperima</i>	1	..	0.4
<i>Dioscorea sp.</i>	3	1.0	2.0	<i>Agave brandegeei</i>	3	0.1	0.3
Gitogenin				<i>Agave cerulata</i>	2	..	Trace
<i>Agave cerulata</i>	1	..	Trace	<i>Agave nelsonii</i>	1	..	0.1
<i>Agave goldmaniana</i>	1	..	Trace	<i>Agave promontorii</i>	1	..	0.1
<i>Agave roseana</i>	2	..	Trace	<i>Agave roseana</i>	4	0.1	0.6
<i>Agave schidigera</i>	1	..	Trace	<i>Agave schidigera</i>	1	..	0.1
<i>Agave aff. schidigera</i>	1	..	0.3	<i>Agave aff. schidigera</i>	1	..	0.7
<i>Agave toumeyana</i>	1	..	Trace	<i>Agave sullivanii</i>	1	..	Trace
<i>Agave virginica</i>	1	..	0.8	<i>Agave sp.</i>	4	Trace	1.6
<i>Agave sp.</i>	2	Trace	0.2	<i>Manfreda sp.</i>	1	..	0.6
<i>Manfreda sp.</i>	2	0.7	1.0	<i>Yucca faxoniana</i>	1	..	0.3
<i>Yucca peninsularis</i>	2	Trace	0.3	<i>Yucca peninsularis</i>	3	0.4	0.9
<i>Yucca whipplei</i>	2	Trace	0.1	<i>Yucca whipplei</i>	2	0.3	0.6
Hecogenin							
<i>Agave atrovirens</i>	1	..	0.8				
<i>Agave aurea</i>	3	0.1	0.7				
<i>Agave brandegeei</i>	3	0.1	0.6				
<i>Agave cerulata</i>	3	Trace	0.2				
<i>Agave chrysantha</i>	1	..	0.2				
<i>Agave datylio</i>	1	..	0.5				
<i>Agave deserti</i>	1	..	0.4				
<i>Agave expansa</i>	1	..	0.1				
<i>Agave goldmaniana</i>	1	..	0.1				
<i>Agave nelsonii</i>	5	0.1	0.7				
<i>Agave parryi</i>	1	..	0.2				
<i>Agave promontorii</i>	2	0.2	0.7				
<i>Agave roseana</i>	3	0.1	0.3				
<i>Agave shawii</i>	1	..	0.2				
<i>Agave sullivanii</i>	1	..	0.1				
<i>Agave toumeyana</i>	1	..	0.6				
<i>Agave vexans</i>	1	..	0.2				
<i>Agave sp.</i>	3	0.2	0.4				
<i>Yucca peninsularis</i>	1	..	0.1				
Manogenin							
<i>Agave aurea</i>	1	..	Trace				
<i>Agave cerulata</i>	2	..	Trace				
<i>Agave chrysantha</i>	1	..	Trace				
<i>Agave goldmaniana</i>	2	..	Trace				
<i>Agave nelsonii</i>	4	Trace	0.7				
<i>Agave promontorii</i>	1	..	Trace				
<i>Agave roseana</i>	2	..	Trace				
<i>Agave shawii</i>	1	..	0.3				
<i>Agave toumeyana</i>	1	..	0.3				
<i>Agave vexans</i>	1	..	0.2				

hecogenin is probably found most frequently, although rarely alone. A few agaves produce smilagenin with the *cis* ring A/B and *iso* carbon 22 configuration. In these cases, the sapogenin is usually not accompanied by isomeric or modified forms. Sarsasapogenin with a *cis* ring A/B and normal carbon 22 configuration was again found as the sole or predominant constituent of a number of western yuccas. However, coastal yuccas, both from the Southwest and Southeast, produce tigogenin and gitogenin. As stated in the previous paper, *Dioscorea* species produce diosgenin as the sole or predominant steroid.

The evidence presented in this and the preceding report, with few exceptions, permits the following generalization. Sapogenin configuration at carbon 5 and carbon 22 is genus and species specific and cannot be altered by the plant or by environmental conditions. Ketonic and hydroxyl groups, however, can be added or removed by numerous species, resulting in the formation of mixtures of sapogenins.

None of the first 1,000 samples gave a triple test for flavonols. In the present list, however, three were found—leaves of *Diervilla richesae* and *Lonicera* sp. in the Caprifoliaceae and leaves and fruit of *Garcinia spicata* in the Hypericaceae. Krewson *et al.* (4), point out that of 27 species of *Eucalyptus*

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examined for rutin, only two contained it. We obtained a test for flavonol in *E. staigeriana* (Myrtaceae), not one of the species hitherto examined. It remains to determine whether this flavonol is rutin.

No test for alkaloid was obtained in 141 samples of *Dioscorea* tubers—18 identified species and 85 lots of unidentified—native to North, Central, and South America, and to the West Indies. In 42 samples native to the rest of the world, alkaloids were found in seven species. This, together with other published evidence, has led to the generalization that the alkaloids probably do not occur in *Dioscorea* in the Western Hemisphere but that they do occur in some Old World species (3). No saponinins were found in *Dioscorea*-containing alkaloids. Of the species which gave a two- or three-plus test for alkaloids, the following are the first on record: *Lycoris squamigera* (Amaryllidaceae); *Buxus harlandi* (Buxaceae); *Lophocereus schottii* (Cactaceae); *Dioscorea dumetorum* (Dioscoreaceae); *Adenocarpus*

foliosus, *Cassia brasiliensis*, *C. emarginata*, *C. excelsa*, *Peltogyne nitens*, *Pithecellobium flexicaule* (Leguminosae); *Neillia longiracemosa* (Rosaceae); *Cephalotaxus henryi* (Taxaceae).

As to tannins, the samples for most families are too few for generalization. However, tannins appear to occur frequently in Anacardiaceae, Caprifoliaceae, Ericaceae, Leguminosae, Myrtaceae, and Rosaceae.

As in the first series, unsaturated sterols were generally abundant and frequent.

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